

UptiReverse Transcriptase 1-Step Kit

Product Description

Name : UptiReverse Transcriptase 1-Step Kit, MgCl₂ FREE

One step system for RT-PCR with thermostable retrotranscriptase

Catalog :	KB0310, 100 tests	KB0312, 500 tests
Contents : UptiReverse transcriptase	55 µl	280 µl
UptiReverse Master Mix	2,8 ml	5 x 2,8 ml
PCR Astringent	1200 µl	5 x 1200 µl
100 mM MgSO ₄ solution	1,8 ml	1,8 ml

Storage: -20°C

Introduction

UptiReverse Transcriptase 1-Step Kit is a novel RT-PCR system for the detection of specific sequences within an RNA sample. This one-step, two-enzyme system uses UptiReverse retrotranscriptase and hot start DNA Polymerase for detection of any type of RNA sample.

The kit has been designed to deliver maximum efficiency, precision and sensitivity of RT-PCR. For these purpose two high-performance enzymes, a thermostable reverse transcriptase and a DNA polymerase with hot start activity, carry out the reactions. Both cDNA synthesis and PCR are performed sequentially in a single tube due to a single buffer system formulated to ensure specific primer annealing.

UptiReverse Transcriptase 1-Step Kit provides a convenient and fast procedure, one step only, for synthesizing cDNA and DNA amplification. All reagents necessary for both reactions are added in one tube at the same time which confers a rapid and easy-to-handle format saving manipulation time and reducing contamination risks without compromising efficiency or sensitivity of the kit. To use it, simply add to a vial containing the prepared Master Mix, desired primers, and RNA template.

Directions for use

Handling and Storage

Store all components of the UptiReverse Transcriptase 1-Step Kit at -20°C in a constant temperature freezer (frost-free freezers are not recommended). All reagents must be thawed and handled on ice. For frequent use, divide in aliquots to avoid multiple freeze-thaw cycles.

- **Master Mix:** Mix before use.
- **PCR Astringent:** Mix before use.
- **MgSO₄ Solution:** Mix thoroughly before use.

If stored under recommended conditions, the product will maintain performance through the indicated date on the label.

Reagents included in the kit

The system contains sufficient reagent for a number of one-tube RT-PCR reactions of 50µl each.

• **Master Mix:** An easy and convenient 2X Master Mix formulated to setting up one-tube RT-PCR. The mixture includes: UptiTherm Split DNA Polymerase, dNTPs, MgSO₄, and Reaction Buffer.

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• **UptiReverse Retrotranscriptase:** A new RNase H minus, thermostable reverse transcriptase which exhibits high affinity for RNA and works in a higher range of temperature than most commercially retrotranscriptases. The RT enzyme is provided in a separate tube.

• **PCR Astringent:** Used for specific RT-PCR assays, to increase sensitivity and specificity of RT-PCRs.

• **100 mM MgSO₄ Solution:** Used only for assays which require an additional optimisation.

General considerations

Template: Successful reverse transcription is dependent on the integrity and purity of template. Samples should be transported and stored frozen. In samples that have been stored without refrigeration, RNA can be degraded.

For optimal results using this system, the RNA template, regardless of the type of RNA using, it should be DNA-free. The polymerase used in this system has no reverse transcriptase activity under standard reaction conditions, but amplification product will be generated out of these reactions if trace amounts of DNA with similar sequences are present in the template preparation.

The presence of carryover of reagents (e.g. SDS, NaCl, heparin, guanidine thiocyanate) from some RNA purification methods can interfere with RT-PCR. We recommend the use of our Speedtools Total RNA Extraction kit.

It is highly recommended determining the concentration by fluorimetry. If you do not know the concentration of template RNA, add a fixed volume of the extraction mixture to problem samples.

The amount of RNA required per reaction depends upon the abundance of the RNA of interest. Up to 1µg RNA can be used in each reaction. A good starting point for a standard mass of RNA to add for an expression level of unknown abundance would be 100ng of total RNA.

MgSO₄ Concentration: The magnesium requirement of both the UptiReverse transcriptase and the Split DNA Polymerase in RT-PCR reactions. The MgSO₄ concentration should be optimised for each experimental target/primer combination. We recommend starting with an initial Mg²⁺ concentration of 6 mM as provided by the 2X High Master Mix. For a few targets, reactions may be improved using Mg²⁺ concentration up to 12 mM. The kit is provided with a vial with 100 mM MgSO₄ for additional optimisation.

PCR Astringent Concentration: The PCR astringent can increase yield of the desired PCR product or decrease production of undesired products. Concentration of astringent has to be optimised for each reaction; the recommended range of volume is 0.8-1.1 µl for RT-PCR reactions of 50µl.

Primer Design: A specific primer should be used for first strand synthesis. Specific primers anneal only to defined sequences and can be used to synthesize cDNA from particular mRNAs rather than from the entire mRNA population in the sample. To differentiate between amplification of cDNA and amplification of contaminating genomic DNA, primers may be designed to anneal to sequences in exons on opposite sides of an intron.

Regardless of primer choice, the final concentration of the primer in the reaction may need to be optimised (between 0.05-0.5µM). We recommend adding 0.2 µM of primer as a starting point of optimisation.

It is recommended to use gene-specific primers designed with a T_m high enough to perform the retrotranscription at 45-47°C.

Synthesis of cDNA: High Scriptools OneStep Kit does not require a template denaturation step prior to initiation of the reverse transcription reaction. If desired, a denaturation step may be incorporated by incubating a separate tube containing primers and RNA template at 95°C for 2 minutes. The template/primers mixture can then be added to the RT-PCR reaction mix for the standard reverse transcription incubation at 45-47°C.

The UptiReverse transcriptase is a thermostable reverse transcriptase working in a wide temperature range, between 40-65°C. We recommend 45-47°C (optimum temperature) to minimised effects of RNA secondary structures and to encourage full-length cDNA synthesis.

Cycling Parameters: Initial denaturation step could be done during 5-10 min at 95°C to denature the RNA/cDNA hybrid; inactivate the UptiReverse transcriptase; and activate the UptiTherm Split DNA Polymerase.

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Using primers with a high T_m may be advantageous to increase the suggested annealing and extension temperatures. The higher temperature minimizes nonspecific primer annealing, thus increasing the amount of specific product produced. For primers with a low T_m , it may be necessary to decrease the annealing temperature to allow the primer to anneal to the target template.

Most RNA samples can be detected using 40 cycles of amplification. If the target RNA is rare or if only a small amount of target is available, it may be necessary to increase the number of cycles to 45-55.

The extension time depends on the size of the expected product; we recommend 1 min for each kb of expected product.

Standard Protocol

Materials to be supplied by user:

- Downstream oligonucleotide primer
- Upstream oligonucleotide primer
- Nuclease-free water

Laboratory workflow must be unidirectional, from pre-amplification to amplification areas. Specific equipment for each working area must be used, in order to avoid cross contaminations. Wear disposable gloves, use nuclease-free material, and filter tips.

KEEP THE REACTION VIALS REFRIGERATED until their introduction in thermal cycler. Be careful not to wet reactions vials.

The inclusion of positive and negative controls is highly recommended in each experiment.

Proceed to the Reagent Preparation Area in a laminar flow cabinet.

- 1.-Thaw and thoroughly mix all reagents before dispensing.
- 2.-Prepare the RT-PCR reaction mix in a sterile 1.5ml microtube on ice as indicated in Table 1. The reaction mix will be used to amplify experimental RNA, NTC and positive control reactions. Prepare sufficient reaction mix for the desired number of reactions on ice.
- 3.-Dispense the appropriate volume of the Reaction Mix in each reaction vial and store vials on ice. The Reaction Mix contains all reagents necessary for the RTPCR except the template.

Table 1. Reaction Mix preparation

COMPONENT	50µl <u>Reaction</u>	Final <u>Concentration</u>
2X MASTER MIX	25µl	1X
MgSO ₄ (100 mM)*	x µl	4-12 mM
Primers	x µl	0,1-0,5 µM
PCR Astringent +	0,8-1,1 µl	-
UptiReverse Transcriptase	0,5 µl	0,5 µl/rxn
Nuclease free water	Up to 50 µl	-
RNA	x µl	Up to <1 µg

* For each experiment the concentration of MgSO₄ >6mM

+ Addition of PCR Astringer depends on your reaction

Proceed to RNA Purification Area.

Never introduce RNA in the laminar flow cabinet from the reagent preparation area. Reaction must start in the next 10 min after adding RNA and primers to the reaction mix. Keep all reagents on ice until their introduction in thermal cycler.

4.-Add template RNA (experimental sample, positive control or NTC) to each reaction tube. Close tubes and mix gently.

5.-Centrifuge amplification vials briefly.

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Proceed to Amplification Area

6.-Place tubes in the thermal cycler and start the RT-PCR cycling program (see Table 2).

Analysis

7.-Analyse PCR products by agarose gel electrophoresis of 10% of the total reaction. Products will be readily visible by UV transillumination of an ethidium bromide-stained gel or other fluorescent intercalating.

8- Store reaction products at -20°C until needed.

Table 2. Cycling parameters for UptiReverse One-Step kit

<u>STEP</u>	<u>TEMP</u>	<u>TIME</u>	<u>COMMENTS</u>
Denaturation	95°C	2 min	<i>Optional : RNA and primer denaturation</i>
RT	42-47°C	30-40 min	<i>Retrotranscription : first strand cDNA synthesis</i>
1 Cycle	95°C	5-10 min	<i>Initial Denaturation and inactivation of retrotranscriptase</i>
PCR*			<i>Second strand cDNA synthesis and PCR</i>
40-50 Cycles	95°C	10-30 sec	<i>Denaturation</i>
	2-5°C < T _m of primers	5-20 sec	<i>Annealing (approximately 5°C below the calculated T_m of the primers)</i>
	60-72°C	45-60 sec	<i>Extension</i>
1 Cycle	72°C	2-5 min	<i>Final extension</i>
	15°C	∞	<i>Cooling</i>

TROUBLESHOOTING

Problem	Cause	Recommendation
Little or no product detected	Check template quality and quantity	Verify the integrity of the RNA by denaturing agarose gel electrophoresis and check the quantity of your template by fluorimetry (an excess RNA can reduce RT-PCR yield). Carryover of reagents from some RNA purification methods can interfere with RT-PCR: reduce volume of target or change purification method. Ensure that reagents, tips and tubes are RNase-free.
	Check primers design and store conditions.	Verify that the downstream primer was designed to be complementary to the downstream sequence of RNA. Design primers that have higher T _m and do not form hairpin loops or primer-dimers. Ensure that storage conditions are adequate
	Optimise primer concentration	Although lower primer concentration can prevent primer-dimer formation, sufficient primers are needed for successful RT-PCR. Increase primer in increments of 0.1 μM.
	Suboptimal reaction conditions	-Optimise the concentration of MgSO ₄ by doing a curve. -Decrease PCR Astringent concentration
	Optimise retrotranscription conditions	Low abundance targets, and/or templates rich in G+C content or with secondary structures often require longer retrotranscription: Increase time up to 60 min. Take care with inactivation of the UptiReverse transcriptase: If an initial denaturation/annealing step is included in the protocol, be certain to add the retrotranscriptase after the denaturation step.
	Optimise PCR Cycling parameters	- <i>Increase the length of initial template denaturation up to 10 min.</i> Templates rich in G+C content or with secondary structures often require a longer step. - <i>Reduce the annealing temperature</i> in 2°C decrements. - <i>Increase N° of cycles</i> in increments of 5 cycles. - <i>Increase extension time</i> by increments of 30 sec.
	Thermal cycler programmed incorrectly	Verify that times and temperature are correct.
	Missing reaction component	Check reaction components, and repeat the reaction.

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Problem	Cause	Recommendation
Multiple, non specific amplification products	Check template quality and quantity	Verify the integrity of the RNA by denaturing agarose gel electrophoresis and check the quantity of your template by fluorimetry. Decrease the amount of RNA and/or primer added to the reaction. If genomic DNA is detected, first treat the template RNA with DNase I, or if this is not possible, redesign primers to avoid amplification of gDNA.
	Check the design and quality of primers	Design primers that have higher T _m and do not form hairpin loops or primer dimers. Check quality of primers by electrophoresis in a denaturing acrylamide gel.
	Increase Retrotranscription temperature	Increase the retrotranscription temperature in 1°C increments.
	Suboptimal reaction conditions	-Increase PCR Astringent concentration -Decrease MgSO ₄ concentration in the reaction mix.
	Optimise PCR cycling parameters	-Increase the annealing temperature in 2°C increments. -Decrease N° of cycles in decrements of 5 cycles.
	Contamination of some reagent	If you view band in negative controls, repeat the assay with new reagents.

References

- New product

Related / associated products and documents

See [BioSciences Innovations catalogue](#) and [e-search tool](#).

- RNase Inhibitor, [973990](#)
- Water, nuclease free, [457420](#)
- pLUG-Multi TA-cloning vector, [MP3370](#)

Ordering information

Catalog size quantities and prices may be found at <http://www.interchim.com>.
Please inquire for higher quantities (availability, shipment conditions).

For any information, please ask : Uptima / Interchim; Hotline : +33(0)4 70 03 73 06

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