DATA SHEET





3'-End Oligonucleotide Labeling Reagent Kit

| Cat. No. | Amount |
|----------|------------------------------------|
| APP-003 | 25 reactions x 50 μl (5 pmol each) |

For research use only!

Shipping: shipped on blue ice

Storage Conditions: store at -20 °C

Additional Storage Conditions: avoid freeze/thaw cycles

Shelf Life: 12 months

Description:

3'-End Oligonucleotide Labeling Reagent Kit contains all buffer reagents required for efficient 3'-End Labeling of DNA oligonucleotides (length: 20 -100 bp, 5 pmol per reaction) except of oligonucleotide template to be labeled and labeled nucleotides..

The labeling principle is based on Terminal deoxynucleotidyl Transferase (TdT) that template-independently transfers labeled nucleotides to the 3'-OH group of ssDNA (e.g. an oligonucleotide) in the presence of CoCl₂. The number of nucleotide and thus label incorporation depends on the type of nucleotide (UTP/ddUTP) and type of label.

Labeled UTP: 1 - 3 label (average)

Labeled dUTP: multiple label (tail length is highly nucleotide specific)

Labeled ddUTPs: 1 label

The resulting 3'-End labeled oligonucleotides are ideally suited for applications involving sequence-specific protein binding or hybridiziation such as EMSA, Northern or Southern blots. Compared to internal, random labeled probes, the label is located at the 3'-End only and less likely interferes with probe binding.

TdT possesses a preference for single-stranded DNA (ssDNA) over dsDNA with 3'-overhangs or blunt ends. For the preparation of labeled dsDNA complexes, label each complementary oligonucleotide separately and anneal them before use.

Content:

Terminal Deoxynucleotidyl Transferase (TdT)

30 μ l (20 U/ μ l) in 100 mM potassium acetate (pH 6.8), 2 mM 2-mercaptoethanol, 0.01% Triton X-100 (v/v) and 50% glycerol (v/v)

5x TdT Reaction Buffer

400 μl containing 1 M potassium cacodylate, 0.125 M Tris, 0.05% Triton X-100 (v/v), 5 mM CoCl $_2$, pH 7.2

Unlabeled Control Oligonucleotide (60 bp)

250 μl, 1 μM in 1x TE Buffer, pH 7.6

PCR-grade H₂O

12.5 ml

1x TE Buffer, pH 7.6

100 ml containing 10 mM Tris-HCl, 1 mM EDTA, pH 7.6

Stop Buffer

400 μl, 0.5 M EDTA solution, pH 8



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3'-End Oligonucleotide Labeling Reagent Kit

1. 3' End Oligonucleotide labeling reaction

- · Store all components except of TdT on ice until use.
- Store TdT at -20°C until use.
- Final Assay volume: 50 μl
- Template requirements: oligonucleotide/ssDNA purified by HPLC or gel electrophoresis, 20 – 100 bp
- · Add all components on ice exactly in the order listed below.
- Mix reaction gently by pipetting up and down. Do not voretex!
- Incubate 30 min at 37 °C.
- Add 1 µl Stop Buffer (0.5 M EDTA solution, pH 8) to stop each reaction.
- Store reactions on ice for subsequent use (see 3.) or -20 °C for long-term storage.

| Component | Volume | Final con- centration | Final molar amount |
|---|---------|--------------------------|-----------------------|
| PCR grade H ₂ O | 31.5 µl | n/a | n/a |
| 5x TdT Reac- tion Buffer | 10 μl | 1x | n/a |
| oligo- nucleotide template (1 µM) | 5 μl | 100 nM | 5 pmol |
| Labeled UTP or ddUTP(10 µM) (see 1.2) | 2.5 μl | 0.5 μΜ | 50 pmol |
| TdT (20 U/μl) | 1 μl | 0.4 U/μl | 20 U |
| Total volume | 50 μl | | |

2. Estimation of Biotin labeling degree

Quantification of labeling degree is essential for reproducible downstream results.

Biotin or Digoxigenin-labeled oligonucleotides can be indirectly detected via Streptavidin or anti-Digoxigenin conjugates, respectively. The labeling degree of fluorescent oligonucleotides can be directly detected by measurement of the nucleic acid-dye conjugate absorbance followed by a calculation of dye to base ratio according to the law of Lambert-Beer.