

**3D Culture BME Cell
Proliferation Assay**

Catalog # 3445-096-K

Table of Contents

**3D Culture BME Cell Proliferation
Assay**

**Reagent kit for investigating cell
proliferation in 3D Culture**

96 samples

	Page
I. Quick Reference Procedure	1
II. Background	2
III. Precautions and Limitations	2
IV. Materials Supplied	2
V. Materials/Equipment Required But Not Supplied	2
VI. Assay Protocol	
A. Cell Harvesting	3
B. Standard Curve	4
C. Cell Proliferation Assays	7
VII. Data Interpretation	9
VIII. Troubleshooting	12
IX. References	12
X. Appendix: Reagent and Buffer Composition	13
XI. Related Products Available From Trevigen	13

Catalog #: 3445-096-K

I. Quick Reference Procedure for Trevigen's 3D Culture BME Cell Proliferation Assay (including the Cell Toxicity Protocol; also see the Tumorigenesis protocol in section VI):

Read through the complete Instructions for Use prior to using this kit. Determine the optimal seeding density for each cell line used. In general, 5,000 cells per well (25,000 cells/cm²) in 100 µl cell culture medium is a good starting point.

1. Culture cells per manufacturer's recommendation; adherent cells should be cultured to no more than 80% confluence.
2. Thaw BME on ice overnight in a 4°C refrigerator.
3. Working on ice, coat each well of the 96 well plate with 35 µl of 3D Culture BME. Centrifuge plates at 300 x g at 4°C for 10 minutes in a swinging bucket rotor to eliminate bubbles and evenly disperse BME across well bottoms.
4. Transfer plate to a CO₂ incubator set at 37°C for one hour to promote gel formation.
5. After one hour, harvest and count cells.
6. Centrifuge cells at 200 x g for 3 min, remove supernatant, and wash with ice cold PBS. Dilute cells to desired concentration in pre-warmed cell culture medium containing 2% BME.
7. Add 100 µl of cells (in 2% BME) per well on top of the gel plug.
8. Incubate at 37°C in CO₂ incubator for 48 - 96 hours.
9. Assay remaining cells for a standard curve (section VI. B.); each cell type will require a separate standard curve.
10. Dilute test compounds to desired concentration in pre-warmed (37°C) cell culture medium, and add 100 µl per well.
11. Add 15 µl of cell proliferation reagent per well, and incubate at 37°C in a CO₂ incubator.
12. Remove 96 well plate, and read absorbance at 450 or 490 nm. Readings may be taken between 1 to 4 hours after the addition of reagent.

II. Background

Recent studies indicate that the composition of the extracellular environment influences cellular responses to apoptosis inducing agents^{1,2} implicating a role for extracellular proteins in influencing both toxicity and drug resistance. As a result, this environment must be mimicked during the course of cell-based studies to provide the most accurate translation to animal models. **Cultrex® Cell Proliferation Assays** were created in an effort to provide more physiologically relevant assessment when using cell models in the screening process for compounds that influence toxicity, cell survival, tumorigenicity, and new tumor formation. These assays offer a flexible, standardized, high-throughput format for quantitating the degree to which pharmacological compounds influence toxicity or tumorigenicity in an *in vivo*-like environment. The **Cultrex® Cell Proliferation Assay** has been adapted to multiple formats so that cell proliferation may be evaluated against different extracellular matrices; the assay is available in the following formats:

- Basement Membrane Extract (BME)
- Laminin I
- Collagen I
- Matrix of Choice

III. Precautions and Limitations

1. For Research Use Only. Not for use in diagnostic procedures.
2. The physical, chemical, and toxicological properties of these products may not yet have been fully investigated; therefore, Trevigen recommends the use of gloves, lab coats, and eye protection while using these chemical reagents. Trevigen assumes no liability for damage resulting from handling or contact with these products.
3. **CULTREX® Cell Proliferation Assays** contain reagents that may be harmful if swallowed, or come in contact with skin or eyes. In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. Material safety data sheets are available on request.

IV. Materials Supplied

<u>Component</u>	<u>Quantity</u>	<u>Storage</u>	<u>Catalog#</u>
3D BME	5 ml	≤-20°C*	3445-005-01
Cell Proliferation Reagent	3 ml	-20°C	3445-096-02
3D Culture 96 Stripwell Plates	2 plates	Room Temp	3445-096-03

*Store in a manual defrost freezer; for long term storage, store at -80°C.

V. Materials/Equipment Required But Not Supplied

Equipment

1. 1 - 20 µl pipettor, 20 - 200 µl pipettor, and 200 - 1000 µl pipettor
2. Laminar flow hood or clean room
3. 37°C CO₂ incubator
4. Low speed swinging bucket 4°C centrifuge and tubes for cell harvesting

- Hemocytometer or other means to count cells
- 20°C storage
- ice bucket
- standard light microscope (or inverted)
- pipette aid
- timer
- 96-well plate reader (450 nm or 490 nm)
- Computer and graphing software, such as Microsoft® Excel®.

Reagents

- Cell Harvesting Buffer; EDTA, trypsin, or other cell detachment buffer.
- Tissue Culture Growth Media, as recommended by cell supplier.
- Pharmacological agents for addition to culture medium, if necessary.
- Sterile PBS or HBSS to wash cells.
- Trypan blue or equivalent viability stain

Disposables

- Cell culture flask, 25 cm² or 75 cm²
- 50 ml tubes
- 1 - 200 µl and 200 - 1000 µl pipette tips
- 1, 5 and 10 ml serological pipettes
- gloves

VI. Assay Protocol

These procedures should be performed in a biological hood utilizing aseptic technique to prevent contamination.

A. Cell Harvesting

Culture cells per manufacturer's recommendation. The following procedure is suggested and may need to be optimized to suit the cell type(s) being studied.

- Cells should be passaged 2 or 3 times prior to use in the assay, and adherent cells need to be 80% confluent. Each well requires approximately 5,000 cells. 25 and 75 cm² flasks yield at least 1 x 10⁶ and 3 x 10⁶ cells, respectively. Determine the number of cells needed to perform a standard curve for each cell type (Section VI.B) and the cell proliferation assay (Section VI. C).
- Prior to harvest, visually inspect cells, and record cell health, relative number, and morphology.
- Wash cells two times with sterile PBS or HBSS. Use 5 ml per wash for a 25 cm² flask and 10 ml per wash for a 75 cm² flask.
- Harvest cells. For 25 cm² flask or 75 cm² flask, add 1 ml or 2 ml, respectively, of Cell Harvesting Buffer (see Materials Required But Not

Supplied), and incubate at 37°C for 5 to 15 minutes until cells have dissociated from bottom of flask.

- Transfer cells to a 15 ml conical tube, and add 5 ml of cell culture medium.
- Centrifuge cells at 200 x g for 10 minutes to pellet cells, remove medium, and resuspend cells in 2 ml of cell culture medium. Cells may need to be gently pipetted up and down with serological pipette to resuspend cells.
- Count cells, and dilute to 1 x 10⁶ cells per ml in cell culture medium.

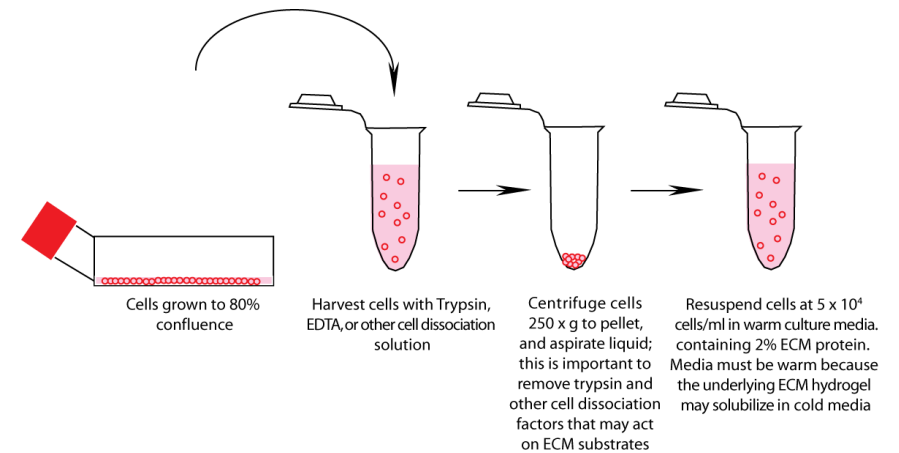


Figure 1. Harvesting and resuspending cells for 3D culture.

B. Standard Curve

The standard curve is necessary to translate OD 450/490 nm to number of cells. A separate standard curve should be run for each cell type, and conditions should be performed in triplicate.

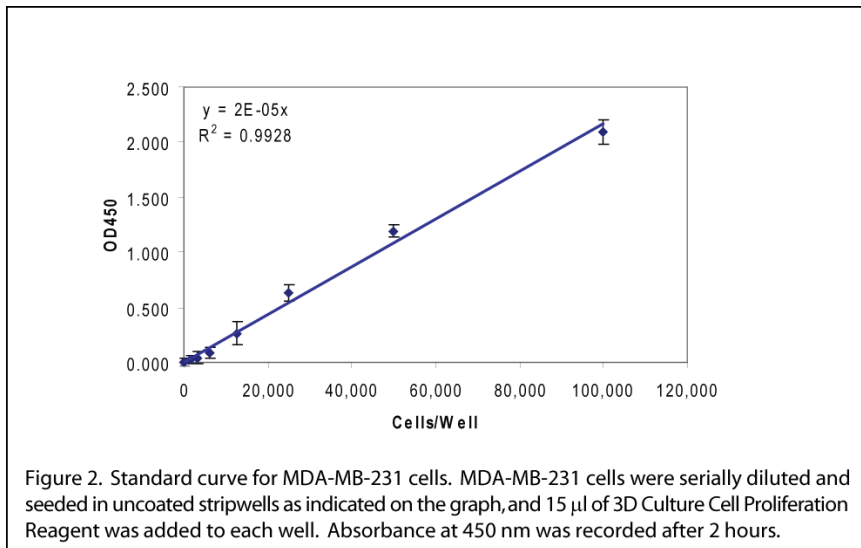
- Determine maximum range of standard curve (eg. 100,000 cells), and develop conditions for standard curve (eg. 100,000 cells, 50,000 cells, 25,000 cells, 10,000 cells, 5,000 cells, 2,000 cells, 1,000 cells, 500 cells, and 0 cells).
- Determine the total number of cells needed for standard curve:

$$= \text{cells/well} \times \text{wells/condition} = \text{cells/condition}$$

$$\text{sum}(\text{cells/condition}) = \text{total number of cells needed}$$
- Calculate the volume of harvested cells needed:

$$= \text{Total number of cells needed} / 1 \times 10^6 \text{ cells/ml}$$
- Transfer volume of harvested cells needed to a 15 ml conical tube, and centrifuge at 200 x g for 10 minutes to pellet cells.
- Remove supernatant, and resuspend cells in culture medium at 1 x 10⁶ cells/ml.

6. Dilute cells to highest condition for a final volume of 200 μl (eg. 100,000 cells/ 200 μl = 5.0×10^5 cells/ml) with cell culture medium. Add 200 μl /well, and serially dilute remaining stock with cell culture medium to generate the desired number of cells per well (in 200 μl of cell culture medium). Repeat dilutions until all conditions have been satisfied. Omit cells from at least three wells to calculate background.
7. Add 25 μL of Cell Proliferation Reagent to each well, and incubate at 37°C, 5% CO_2 .
8. Read absorbance of plate at 450 nm or 490 nm at incubation periods of 1, 2, 3, and four hours (see Table 1 for sample data).
9. Average values for each condition; then subtract background from each value (see Table 2).
10. Plot standard curve of absorbance vs. number of cells (see Figure 2).
11. Insert a linear trendline (best fit) with y intercept at zero, Determine optimal incubation period based on minimum standard deviations and maximum R^2 values.
12. Use the line equation for each cell line to calculate number of cells in each well.



Sample data for standard curve:

Table 1. Raw data for MDA-MB-231 standard curve.

Cells/Well	OD 450 nm			
100.000	2.211	2.362	2.458	2.432
50.000	1.457	1.520	1.477	1.397
25.000	1.006	0.833	0.873	0.890
12.500	0.581	0.583	0.375	0.593
6.250	0.414	0.367	0.319	0.319
3.125	0.382	0.309	0.297	0.243
1.563	0.337	0.291	0.281	0.260
0	0.316	0.276	0.216	0.265

Table 2. Values corrected for background.

Cells/Well	OD 450 nm (corrected)			
100.000	1.942	2.093	2.190	2.164
50.000	1.188	1.252	1.208	1.129
25.000	0.737	0.565	0.604	0.621
12.500	0.313	0.315	0.107	0.325
6.250	0.146	0.098	0.050	0.051
3.125	0.113	0.041	0.029	-0.026
1.563	0.068	0.022	0.013	-0.009
0	0.048	0.008	-0.052	-0.003

Table 3. Summary of averages for corrected values for MDA-MB-231 standard curve.

Cells/Well	Average	Std Dev
100.000	2.097	0.111
50.000	1.194	0.051
25.000	0.632	0.074
12.500	0.265	0.105
6.250	0.086	0.046
3.125	0.039	0.057
1.563	0.024	0.032
0	0.000	0.041

C. Cell Proliferation Assay

Prior to Day 1

1. Determine optimal seeding density for each cell line used. In general, 5,000 cells per well (25,000 cells/cm²) in 100 µl cell culture medium is a good starting point.
2. Culture cells per manufacturer's recommendation; adherent cells should be cultured to no greater than 80% confluence.
3. Thaw BME on ice overnight in a 4°C refrigerator.

Day 1

Coat Stripwells

4. Working on ice, coat each well of the 96 well plate with 35 µl of 3D Culture BME. Centrifuging plates at 200 x g at 4°C for 10 minutes in a swinging bucket rotor will eliminate any bubbles resulting from pipetting and disperse the coating evenly across the bottom of the wells.
5. Transfer plate to a CO₂ incubator set at 37°C for one hour to promote gel formation.

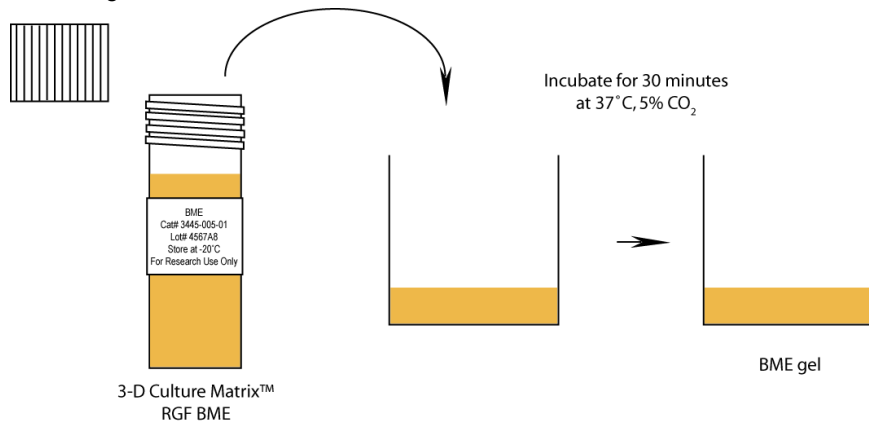


Figure 3. Coating stripwells with BME.

6. After one hour, harvest and count cells, as directed in section VI, A.
7. Centrifuge cells at 200 x g for 3 min, remove supernatant, and wash with ice cold PBS; then dilute cells to the desired concentration in cell culture medium containing 2% BME warmed to 37°C.
8. Add 100 µl of cells to each stripwell.
9. Assay remaining cells for standard curve (section VI, B); each cell type will require a separate standard curve.
10. Continue to either Toxicity Assay or *in vitro* Tumorigenicity Assay.

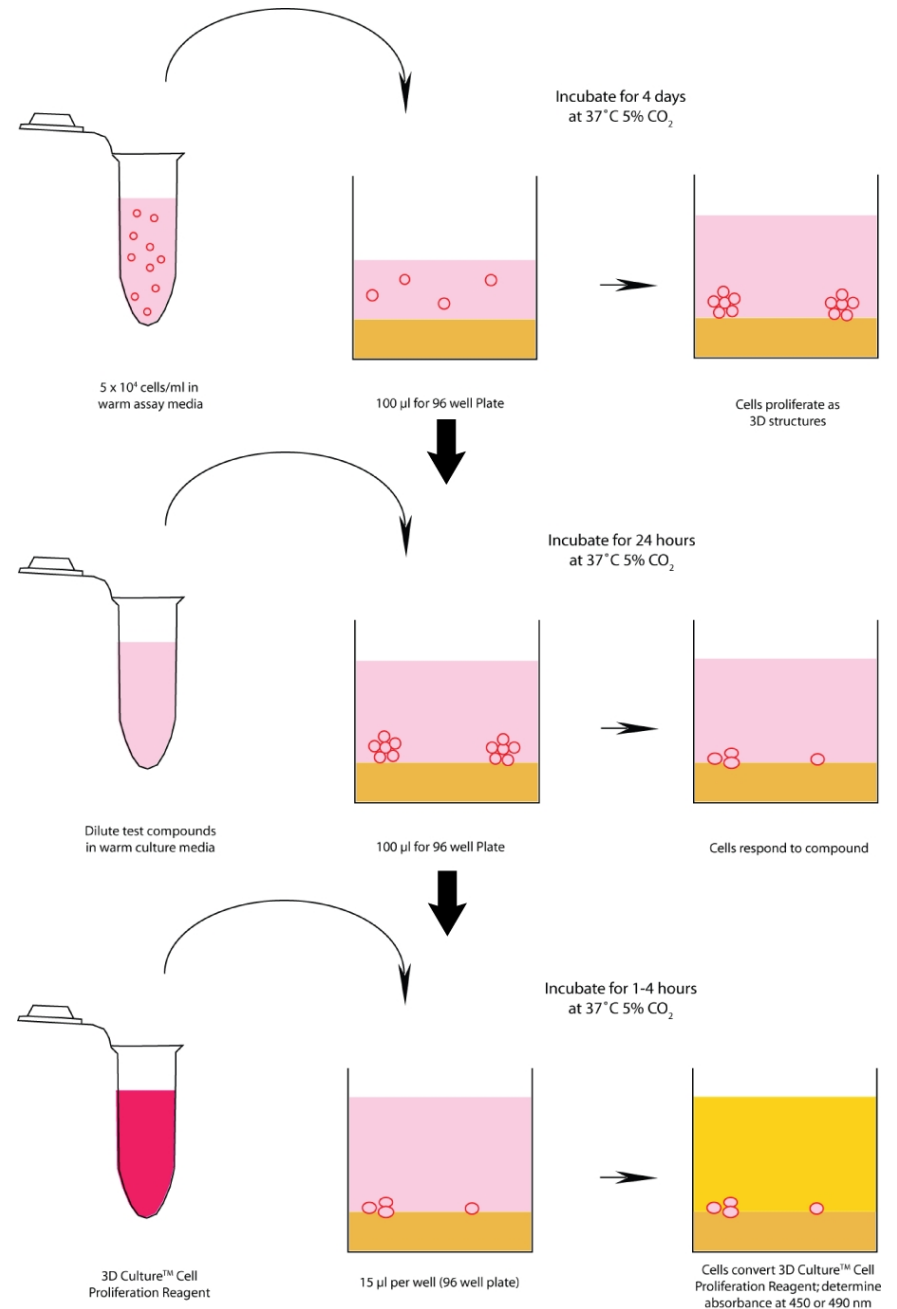


Figure 4. Illustration of protocol for assessment of toxicity in 3D culture.

3D Culture Toxicity Assay:

This assay tests the ability of compounds to induce cell death after the establishment of 3D structures.

1. Culture cells at 37°C in CO₂ incubator for 48 - 96 hours to promote the formation of 3D structures (Figure 4).
2. When structures have reached desired size/morphology, dilute test compound(s) in cell culture medium.
3. Add 100 µl of media containing test compound to each well, and incubate at 37°C in CO₂ incubator for 24 hours.
4. Once incubation is complete, add 15 µl of 3D Culture™ Cell Proliferation Reagent per well.
5. Assess absorbance at 450 or 490 nm between 1-4 hours. Optimal incubation times should be determined when performing the standard curve, based on linearity of curve (lowest R²) accompanied by the lowest standard deviations.

In Vitro Tumorigenicity/Toxicity Assay:

In the body, metastasizing cells can enter the circulation and emerge as single cells that can establish tumors. This assay tests for the inhibition of proliferation and formation of 3D structures, starting from single cells.

1. Dilute test compounds as desired in warm (37°C) cell culture medium, and add 100 µl per well.
2. Incubate at 37°C in CO₂ incubator for 48 - 96 hours.
3. Once incubation is complete, add 15 µl of 3D Culture™ Cell Proliferation Reagent per well.
4. Assess absorbance at 450 or 490 nm between 1-4 hours. Optimal incubation times should be determined when performing the standard curve, based on linearity of curve (lowest R²) accompanied by the lowest standard deviations.

VII. Data Interpretation

1. After plotting a standard curve (section VI, B) with Y intercept at zero, insert trendline, equation, and R-square value (coefficient of determination), as demonstrated in Figure 2.
2. For assay samples, first average all wells for each condition, in the same fashion as was done for the standard curve, Table 1.
3. Next, subtract background from averages, similar to Table 2.
4. Use the trendline equation from the standard curve to determine the number of cells present in each well; for the equation, $y = mx + b$, replace Y value with OD 450 or 490 nm, and solve for X. See an example of a trendline and equation in Figure 2.
5. For each test sample, the number of cells may be compared to the number of cells present in the untreated control.

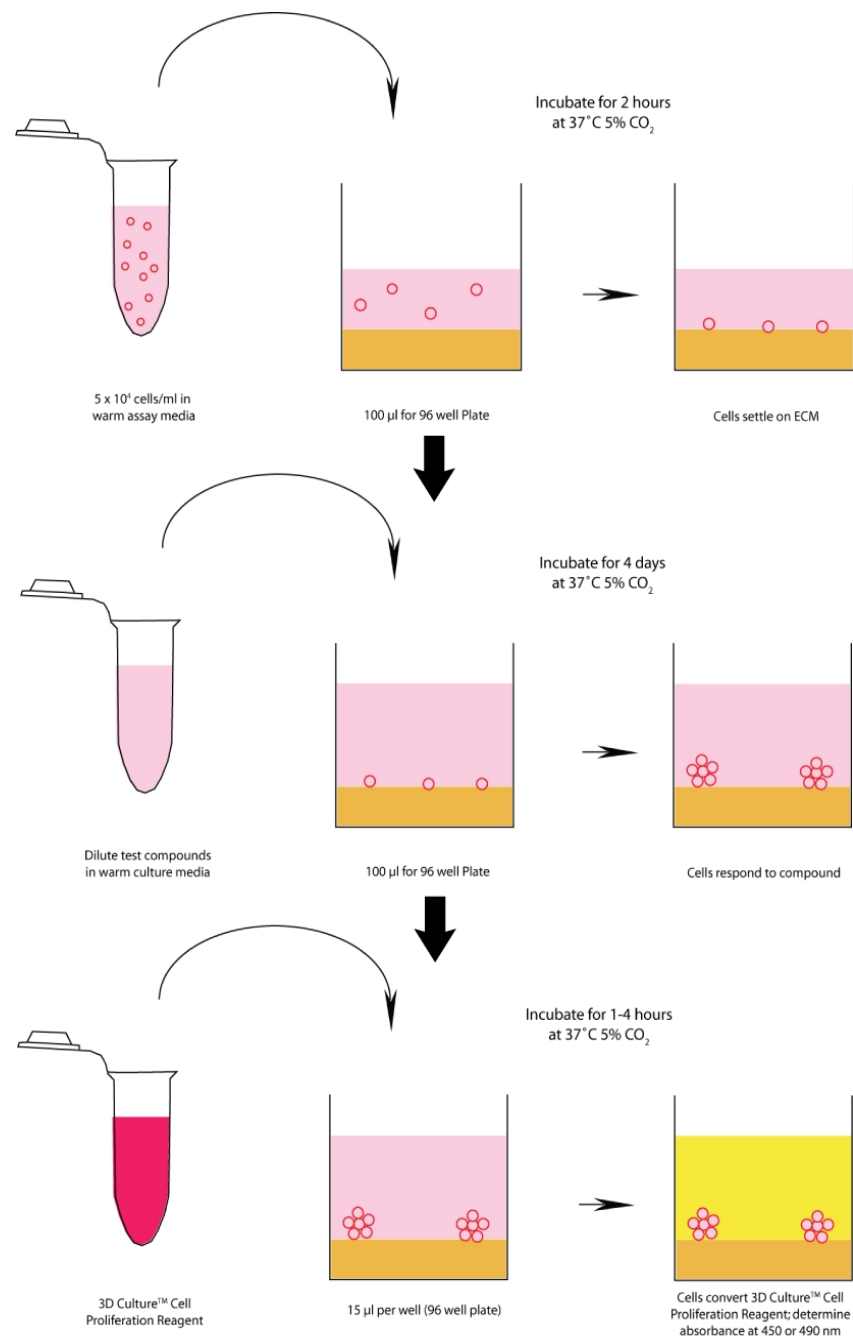
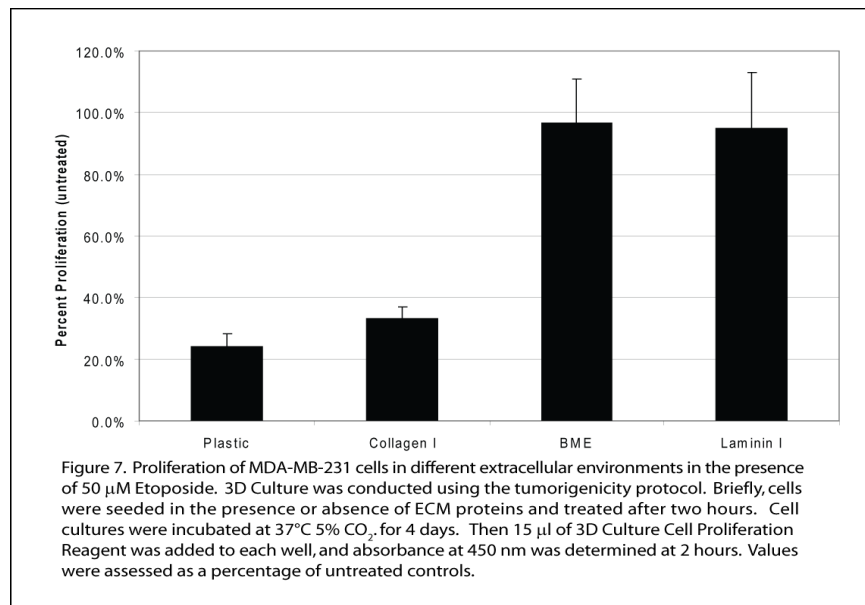
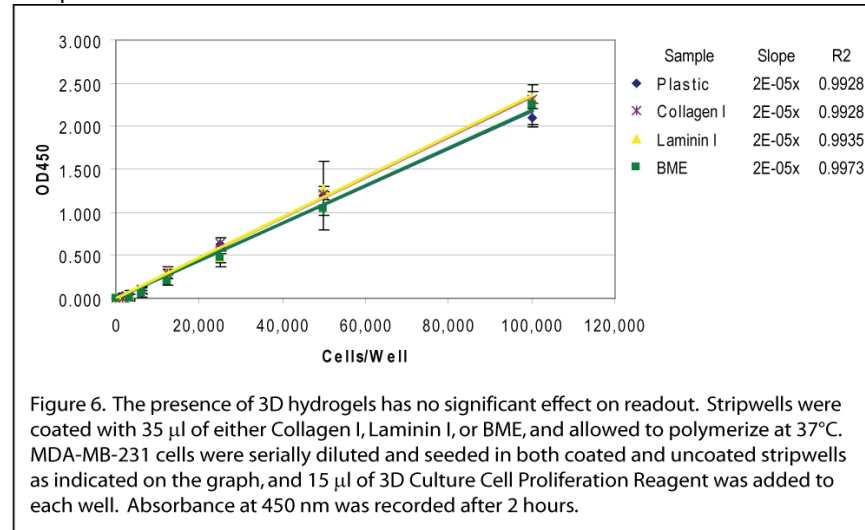


Figure 5. Illustration of protocol for assessment of tumorigenicity in 3D culture.

- Figure 6 demonstrates that there is no significant effect of 3D hydrogels on readout for this assay.
- Figure 7 demonstrates the practical application of this assay in determining ECM-modulated resistance of an apoptosis inducing reagent, etoposide.

Sample Data:



VIII. Troubleshooting

Problem	Cause	Solution
Low/No signal	Instrument not set up properly.	Read absorbance at 450 or 490 nm; adjust gain for optimal sensitivity, if applicable.
	Insufficient cell number	Increase cell number.
	Insufficient incubation period with substrate	Increase incubation period.
	Cells may have died as a result of treatment.	Test cells for viability in treatment regimen (dose response curve).
High background	Instrument not set up properly.	Read absorbance at 450 or 490 nm; adjust gain for optimal sensitivity, if applicable.
	Contamination - proteases released by bacteria or mold may affect Cell Proliferation Reagent	Start a new culture from seed stocks, and re-assay. If seed stock is contaminated, then it may be prudent to get new cells.
Well to well variability	Inconsistent pipetting	Calibrate pipettors, and monitor pipet tips for air bubbles.
	Poor cell dissociation	Pipet cells up and down to create single cell suspension.

IX. References

- Aoudjit F, Vuori K. (2001) Integrin signaling inhibits paclitaxel-induced apoptosis in breast cancer cells. *Oncogene* 20:4995-5004.
- Hodkinson PS, Elliott T, Wong WS, Rintoul RC, Mackinnon AC, Haslett C, T Sethi. (2006) ECM overrides DNA damage-induced cell cycle arrest and apoptosis in small-cell lung cancer cells through β 1 integrin-dependent activation of PI3-kinase. *Cell Death and Differentiation* 13, 1776–88.

X. Appendix A: Reagent and Buffer Composition

1. 3D Culture BME

Basement membrane extract derived from murine EHS sarcoma cells. The reconstituted basement membrane is primarily composed of Laminin I, Collagen IV, Entactin, and heparin sulfate proteoglycans. It is provided in DMEM with 10 µg/ml gentamycin. Avoid freeze-thaws.

2. Cell Proliferation Reagent

Water soluble tetrazoleum substrate with electron coupler.

3. 96 Stripwell Plates

Clear, TC-treated stripwell plates.

XI. Related product available from Trevigen.

Catalog #	Description	Size
4890-025-K	TACS [®] MTT Cell Proliferation Assay	2500 tests
4891-025-K	TACS [®] XTT Cell Proliferation Assay	2500 tests
4892-010-K	Cultrex [®] Calcein-AM Cell Viability Kit	1000 tests
4817-60-K	FlowTACS [™] Apoptosis Detection Kit	60 samples
4822-96-K	HT TiterTACS [™] Assay Kit	96 tests
4830-01-K	TACS [®] Annexin V FITC Kit	100 samples
4835-01-K	Annexin V Biotin Kit	100 samples
6300-100-K	DePsipher [™] Mitochondrial Potential Assay Kit	100 tests
6305-100-K	MitoShift [™] Mitochondrial Potential Assay Kit	100 tests

The product accompanying this document is intended for research use only and is not intended for diagnostic purposes or for use in humans.

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