

G-DEX™ IIc Genomic DNA Extraction Kit

Representative Protocol For $1\sim 2 \times 10^6$ Cells

Notice Before Use

- See the tables for the best result:
[Table 1] Solution Amounts According to Cell Number
[Table 2] Appropriate Tube Selection Guide
[Table 3] Recommended Cell Number
- Recommended cell number is based on HeLa cell. Please refer [Table 3] and it can be modified for adhesive cells.
- This protocol is for genomic DNA extraction from cultured cell ($1\sim 2 \times 10^6$ cells).

Kit Contents (CAT. NO. 17231)

Lysis Buffer 60ml	PPT Buffer 25ml	RNase A Sol. 300 μ l	DNA Rehyd. Buffer 25ml
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Representative Protocol

This protocol is for extraction of genomic DNA from $1\sim 2 \times 10^6$ cells and the recommended condition can be depended on the number of cells (under-lined). Please, refer the [Table 1].

1st Cell Lysis Step

1. Prepare $1\sim 2 \times 10^6$ cells in PBS into 1.5 ml tube.
2. Centrifuge at 13,000~16000 x g for 5 sec (RT), then remove supernatant except 5~10 μ l.
[Note 1] The remained solution can help to resuspend the pellet easily.
[Note 2] It is recommended to use the tabletop centrifuge at maximum rpm.
3. Resuspend the pellet completely. [Note 3] Leaving no cell clumps.
4. Add 150 μ l of Cell Lysis Buffer into the tube and lysis the cells by pipetting 5-20 times.
[Note 4] If there are some clumps, incubate the tube at 37 $^{\circ}$ C until the clumps are dissolved completely.

2nd RNase Treatment Step

1. Add 1 μ l of RNase A Solution into the tube, and incubate at 37 $^{\circ}$ C for 15~30 min.
[Note 5] It is recommended to mix occasionally during the incubation.
[Note 6] RNase A solution can be added after resolving DNA in DNA Hydration Buffer .

3rd Protein Precipitation Step

1. Cool down until room temperature and add 50 μ l of PPT Buffer, then vortex for 20 sec.
[Note 7] This step is essential to remove protein. Samples can be incubated on ice if necessary.
2. Centrifuge at 13,000~16,000 x g for 3~5 min, this step for the removing the protein pellet.
[Note 8] If the protein pellet is not visible, incubate the tube on ice for 5 min and centrifuge it again.

4th DNA Precipitation Step

1. Transfer the aqueous phase in to a new tube and add 150 μ l of isopropanol (2-propanol, 100%), then invert tubes several times.
[Note 9] If DNA yield poorly, add 1/100 μ l of glycogen(20 mg/ml) before adding isopropanol. It is recommended to adjust aqueous phase volume and 2-propanol in the ratio of 1:1.
2. Centrifuge at 13,000~16,000 x g for 1 min to gain DNA pellet.
3. Discard the supernatant and keep the tube on the paper towel to dry the pellet.
Add 150 μ l of 70% ethanol and invert several times, then centrifuge at 13,000~16,000 x g for 1 min.
[Note 10] 500 μ l ~ 1 ml of 70% ethanol can be used.
4. Discard aqueous phase carefully and the pellet should be air-dried for 10~15 min.
[Note 11] Overdrying the pellet may make the DNA difficult to dissolve again.

5th DNA Rehydration Step

1. Add 150 μ l of DNA Rehydration buffer and dissolve the pellet.
[Note 12] The amount of DNA Rehydration buffer can be adjusted by DNA concentration. Refer the table 1.
2. If the pellet is over-dried, incubate at 65 $^{\circ}$ C for 30~60 min or 4 $^{\circ}$ C for O/N.
3. Extracted DNA concentration and purity (A_{260}/A_{280}) should be determined by UV spectrophotometer.
4. Store at -20 $^{\circ}$ C (short term) or -80 $^{\circ}$ C (long term).

Protocol

Protocol

Table 1 Solution Amounts According to Cell Number (K562)

•We recommend this solution amounts table for the best result.

Cell Number (cells)	Cell Lysis	RNase A	PPT Buffer	Isopropanol (not supplied)	70% EtOH (not supplied)	Rehydration buffer	DNA Yields
100 ~ 10,000	60 µl	0.5 µl	20 µl	60 µl	60 µl	25 µl	43 ± 6 ng
10,000 ~ 100,000	100 µl	0.5 µl	33 µl	100 µl	100 µl	25 µl	45 ~ 910 ng
1.0 ~ 9.0 × 10 ⁵	150 µl	1.0 µl	50 µl	150 µl	150 µl	50 µl	1 ~ 12 µg
1.0 × 10 ⁶	150 µl	1.0 µl	50 µl	150 µl	150 µl	100 µl	13 ± 4 µg
1.0 ~ 2.0 × 10 ⁶	300 µl	1.5 µl	100 µl	300 µl	300 µl	150 µl	9 ~ 17 µg
3.0 ~ 5.0 × 10 ⁶	600 µl	3.0 µl	200 µl	600 µl	600 µl	200 µl	20 ~ 35 µg
5.0 × 10 ⁶	600 µl	3.0 µl	200 µl	600 µl	600 µl	200 µl	35 ± 4 µg
6.0 ~ 9.0 × 10 ⁶	750 µl	4.5 µl	250 µl	750 µl	750 µl	300 µl	33 ~ 44 µg
1.0 × 10 ⁷	1.5 ml	7.5 µl	500 µl	1.5 ml	1.5 ml	500 µl	48 ± 6 µg
1.0 ~ 2.0 × 10 ⁷	3.0 ml	15 µl	1 ml	3.0 ml	3.0 ml	500 µl	46 ~ 134 µg
3.0 ~ 5.0 × 10 ⁷	6.0 ml	30 µl	2 ml	6.0 ml	6.0 ml	600 µl	84 ~ 331 µg
5.0 × 10 ⁷	6.0 ml	30 µl	2 ml	6.0 ml	6.0 ml	600 µl	328 ± 19 µg
6.0 ~ 9.0 × 10 ⁷	10.0 ml	50 µl	3.3 ml	10.0 ml	10.0 ml	800 µl	306 ~ 470 µg
1.0 ~ 2.0 × 10 ⁸	15.0 ml	75 µl	5 ml	15.0 ml	15.0 ml	1 ml	560 ~ 1,700 µg

(1) Isopropanol and 70% ethanol are not supplied.

(2) K562 cell line is used for this table and other cell line may be different slightly.

(3) The amount of isopropanol is based on 90 ~ 95% of aqueous phase. It is recommended to adjust aqueous phase volume and 2-propanol in the ratio of 1:1.

(4) More Rehydration (1.5~3 times) buffer is required to redissolve the pellet. Note that DNA concentration should be considered.

(5) If DNA yield poorly, add 1/100 µl of glycogen (20 mg/ml) before adding isopropanol. Glycogen is a highly purified polysaccharide that can be used as a carrier for nucleic acid PPT.

Table 2

:Appropriate Tube Selection Guide

Cell Number	Tube
100 ~ 10,000	0.6 ml tube
10,000 ~ 5.0 × 10 ⁶	1.5 ml tube
6.0 ~ 9.0 × 10 ⁶	2.0 ml tube
1.0 × 10 ⁷ ~ 5.0 × 10 ⁷	15 ml tube
6.0 × 10 ⁷ ~ 2.0 × 10 ⁸	50 ml tube

There are many kinds of dishes and flasks for the cell culture, so we describe them to the table 3, it can be shown the proper seeding density and confluency depending on the cell number. This table is based on the HeLa Cell.

Table 3 Useful Numbers for Cell Culture

	Surface Area (mm ²)	Seeding Density	Cells at Confluency
Dishes			
35 mm	962	0.3 × 10 ⁶	1.2 × 10 ⁶
60 mm	2,827	0.8 × 10 ⁶	3.2 × 10 ⁶
100 mm	7,854	2.2 × 10 ⁶	8.8 × 10 ⁶
150 mm	17,671	5.0 × 10 ⁶	20.0 × 10 ⁶
Plates			
6-well	962	0.3 × 10 ⁶	1.2 × 10 ⁶
12-well	401	0.1 × 10 ⁶	0.4 × 10 ⁶
24-well	200	0.5 × 10 ⁶	0.2 × 10 ⁶
Flasks			
T-25	2,500	0.7 × 10 ⁶	2.8 × 10 ⁶
T-75	7,500	2.1 × 10 ⁶	8.4 × 10 ⁶
T-160	16,000	4.6 × 10 ⁶	18.4 × 10 ⁶

Related Products

ITEM	CAT. NO.	SIZE
G-spin™ Genomic DNA Kit (For Cell/Tissue)	17041	50 col.
G-spin™ Genomic DNA Kit (For Blood)	17111	50 col.
G-spin™ Genomic DNA Kit (For Bacteria)	17121	50 col.
G-spin™ Genomic DNA Kit (For Plant)	17191	50 col.
Viral Gene-spin™ Viral DNA/RNA Extraction Kit	17151	50 col.
RNA-spin™ Total RNA Extraction Kit	17211	50 col.