Preparation of FISH probe

- 1. The following FISH probes are ready to use, no need of any preparation.
 - a. Gene FISH Probe (Cat # FGxxxx)
 - b. Split FISH Probe (Cat # FSxxxx)
 - c. Translocation FISH Probe (Cat # FTxxxx)
 - d. Prenatal FISH Probe (Cat # FMxxxx)
 - e. Made to Order FISH Probe (Ca # FAxxxx)
- 2. Chromosome FISH Probe (Cat # FCxxxx) and Subtelomere FISH Probe (Cat # FExxxx) are provided in 5x concentrated format, they should be either:
 - a. Diluted to 1x with FISH Hybridization Buffer (Cat # <u>U0028</u> or <u>U0029</u>) before use,

OR

b. Mixed with same category of FISH Probes (up to 5 different probes) to use, for example:

Combine 2 different probes:

1 volume of probe 1 (2 uL) + 1 volume of probe 2 (2 uL)

+ 3 volume of FISH Hybridization Buffer (6 uL)

Combine 3 different probes:

1 volume of probe 1 (2 uL) + 1 volume of probe 2 (2 uL)

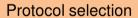
- + 1 volume of probe 3 (2 uL)
- + 2 volume of FISH Hybridization Buffer (4 uL)

Recommended filter set

The table below is a recommendation of filter set use:

| Fluorophore | Brand | Recommended filter set |
|--------------------------------|---------|------------------------|
| Single fluorophore: | | |
| FITC (EX. 426; EM. 480) | Semrock | SpGr-B |
| Texas Red (EX. 593; EM. 612) | Semrock | SpRed-B |
| DEAC (EX. 426; EM. 480) | Semrock | SpAqua-C |
| R6G (EX. 525; EM. 550) | Semrock | SpGold-B |
| Cy5 (EX. 650; EM. 668) | Semrock | CY5-4040B |
| | | |
| Multiple fluorophores: | | |
| FITC, Texas Red & DAPI | Semrock | DA/SpGr/SpRed-A |

Note: EX. = excitation wavelength; EM. = emission wavelength



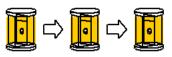
Please follow an appropriate protocol below depend on the sample use, these samples include **Paraffin embedded tissue (or FFPE)**, **Frozen tissue** and **Metaphase spreads**.

For Paraffin embedded tissue, we recommended FFPE FISH PreTreatment Kit 1(Catalog #: <u>KA2375</u> or <u>KA2691</u> for the pretreatment of Formalin-Fixed Paraffin-Embedded (FFPE) tissue sections.



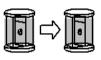
Paraffin embedded tissue

1. Deparaffinized



Xylene 5minx3 Room temperature

2. Dehydrate

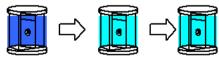


100%EtOH 5minx2 Room temperature

3. Air dry



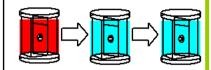
4. Pre-treatment



Paraffin
Pretreatment
Solution
95° 30min

Wash buffer (2×SSC) 5min × 2

5. Protease treatment



Wash buffer (2×SSC) 5min × 2

 $\mbox{\begin{tabular}{l} \curvearrowright}$ Protease Solution Add 500 μ I protease in 50ml protease buffer

6. Dehydrate (Room temperature)







70% EtOH 1min

100% EtOH 1min

7. Air dry

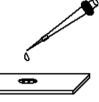


FISH protocol

1. Mark hybridizing area



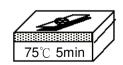
2. Apply 10µl FISH probe for 22mm x 22mm area



3. Cover with cover glass Seal with rubber cement



4. Denature

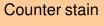


Hybridization

1. Incubation



Humidified box 37° C $16 \sim 72$ hrs



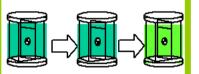
1. Apply 10µl DAPI Solution to target area



☆DAPI Paraffin embedded tissue 1500ng/ml

Wash procedure

Remove rubber cement Slide into 2X SSC and remove cover glass



2X SSC 2X SSC 2X SSC Room temp. /0.3% NP-40 Room temp 5min 73~75°C 1min 1-2min 2. Put on cover glass Seal with manicure









Page 2/4 (Ver. 1.6) GSP Lab., Inc.

Frozen tissue

- 1. Frozen tumour tissue
- 2. Air dry



Positive charged slides

3. Fix and Dehydrated

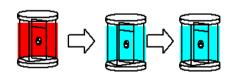


95%EtOH 20min

4. Air dry



5. Protease treatment



Protease Solution 37°C 10∼20min

Wash buffer (2×SSC) 5min × 2

 ${\rm \begin{tabular}{l} \nearrow}$ Protease Solution Add 50 μ I protease in protease buffer

 6. Dehydrate (Room temperature)







70% EtOH 1min

100% EtOH 1min

7. Air dry



touch preparations of unfixed

tumourtissue/cell smears/cytospins

of culturedor blood cells are possible

FISH protocol

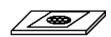
1. Mark hybridizing area



2. Apply 10µl FISH probe for 22mm x 22mm area



3. Cover with cover glass Seal with rubber cement



4. Denature



Hybridization

1. Incubation



Humidified box 37° C $16 \sim 72 \text{ hrs}$

Counter stain

1. Apply 10µl DAPI Solution to target area



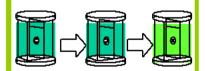
☆DAPI Frozen tumour tissue 150ng/ml

2. Put on cover glass Seal with manicure



Remove rubber cement Slide into 2X SSC and remove cover glass

Wash procedure



2X SSC 2X SSC 2X SSC Room temp. /0.3% NP-40 Room temp. 5min 73~75°C 1min 1-2min







Page 3/4 (Ver. 1.6) GSP Lab., Inc.

Metaphase spreads

1. Ageing



37°C 30min

Ageing solution (2XSSC/0.1% NP-40:PH7~8)

| 20X SSC | 5ml |
|---------|------|
| NP-40 | 50µl |
| DDW | 45ml |

2. Dehydrate (Room temperature)







70% EtOH 1min 100% EtOH 1min

3. Air dry



FISH protocol

1. Slide preparation



73~75°C 5min

<u>Denaturant Solution</u> (2XSSC/70%formamide : PH7~8)

| 100%formamide | 35ml |
|---------------|------|
| 20XSSC | 5ml |
| DDW | 10ml |
| ` | |

2. Dehydrate

(Room temperature)







70% EtOH 1min 100% EtOH 1min

3. Air dry



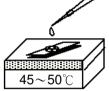
Probe preparation



10 μ l 73 \sim 75 $^{\circ}$ C 5min

Hybridization

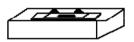
1. Apply 10µl FISH probe for 22mm x 22mm area



2. Cover with cover glass Seal with rubber cement



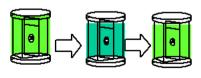
3. Incubation



Humidified box 37°C 16 ~ 72 hrs

Wash procedure

Remove rubber cement Slide into 2X SSC and remove cover glass



2X SSC Room temp. 5min

0.4X SSC /0.3% NP-40 73~75°C 1-2min

2X SSC Room temp. 1min

Counter stain

1. Apply 10µl DAPl Solution to target area



☆DAPI Metaphase spreads 150ng/ml

2. Put on cover glass Seal with manicure







