



## FavorPrep™ Viral Nucleic Acid Extraction Kit II

- For isolation of viral nucleic acid from cell-free fluid such as, serum, plasma, body fluid and cell cultured supernatant

Cat.No. : FVNKK 000-2, 4 Preps  
FAVNK 002, 50 Preps  
FAVNK 002-1, 100 Preps  
FAVNK 002-2, 300 Preps

### Kit Contents:

(For Research Use Only)

	FAVNK 000-2 (4 preps_sample)	FAVNK 002 (50 preps)	FAVNK 002-1 (100 preps)	FAVNK 002-2 (300 preps)
AD Buffer * (concentrate)	0.4 ml	4 ml	8 ml	24 ml
VNE Buffer	1.8 ml x 2	30 ml	60 ml	180 ml
Wash Buffer 1 * (concentrate)	0.9 ml x 2	22 ml	44 ml	132 ml
Wash Buffer 2 * (concentrate)	1.5 ml	20 ml	20 ml x 2	50 ml x 2
RNase-free Water	0.5 ml	6 ml	12 ml	30 ml
VNE Column	4 pcs	50 pcs	100 pcs	300 pcs
Collection Tube	8 pcs	100 pcs	200 pcs	600 pcs
Elution Tube	4 pcs	50 pcs	100 pcs	300 pcs
User Manual	1	1	1	1

\* Preparation of AD Buffer, Wash Buffer W1 and Wash Buffer W2 for first use:

Cat. No:	FAVNK000-2 (4 preps)	FAVNK002 (50 preps)	FAVNK002-1 (100 preps)	FAVNK 002-2 (300 preps)
ethanol volume for AD Buffer	3 ml	30 ml	60 ml	180 ml
ethanol volume for Wash Buffer 1	0.33 ml	8 ml	16 ml	48 ml
ethanol volume for Wash Buffer 2	6 ml	80 ml	80 ml	200 ml

### Specification:

Principle: spin column (silica membrane)

Sample: 150 µl cell-free fluid such as serum, plasma, body fluid and cell cultured supernatant

Fragment size: 100 bp ~30 kb

Recovery rate: 70 ~ 90 %

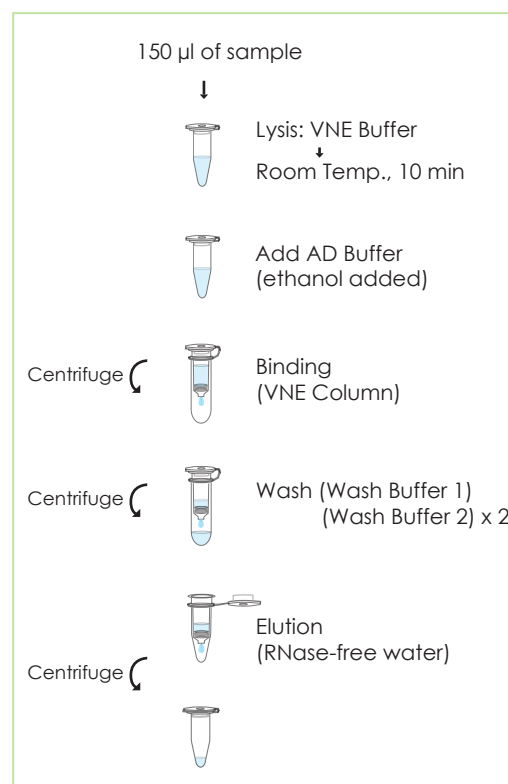
Binding capacity: 30 ug

Elution volume: 40 ~ 50 µl

Operation time: 20 min

### Important Notes:

1. Make sure everything is RNase-free when handling this system.
2. Buffers provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
3. Add required ethanol (96-100%) to AD Buffer, Wash Buffer 1 and Wash Buffer 2 before use.
4. Preheat RNase-free water to 70°C for elution step. (step:10)



## General Protocol:

### Please Read Important Notes Before Starting Following Steps.

HINT: Preheat RNase-free water 70 °C for step 11 (elution step).

- 1. Transfer 200 µl of sample (serum, plasma, body fluids or cell cultured supernatant ) into a microcentrifuge tube (not provided).**  
--If prepared sample is less than 200 µl , adjust sample volume to 200µl with PBS (not provided).
- 2. Add 500 µl of VNE Buffer the sample, mix well by vortexing, and incubate for 10 minutes at room temperature.**
- 3. Add 550 µl of AD Buffer (ethanol added) to the sample mixture and mix well immediately by plus-vortexing.**  
--Make sure that ethanol has been added into AD Buffer when first open.
- 4. Combine a VNE column with a Collection Tube (provided).**
- 5. Transfer up to 750 µl of sample mixture (ethanol added) to the VNE Column, centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.**
- 6. Transfer the rest of sample mixture (ethanol added) to the VNE Column, centrifuge at 8,000 x g for 1 min.**  
Discard the flow-through and the Collection Tube. Combine the VNE Column with a new Collection Tube (provided).
- 7. Add 500 µl of Wash Buffer 1 (ethanol added) to the VNE Column, centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.**  
--Make sure that ethanol (96~100%) has been added into Wash Buffer 1 when first open.
- 8. Add 750 µl of Wash Buffer 2 (ethanol added) to the VNE Column, centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.**  
--Make sure that ethanol (96~100%) has been added into Wash Buffer 2 when first open.
- 9. Repeat step 8. Add 750 µl of Wash Buffer 2 (ethanol added) to VNE Column, centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.**
- 10. Centrifuge at full speed (~18,000 X g) for an additional 3 min to dry the VNE column. Discard the flow-through and the Collection Tube.**  
--**Important step!** This step will avoid the residual liquid to inhibit the subsequent enzymatic reactions.
- 11. Combine the VNE Column with a Elution Tube (provided). Add 50 µl of preheated RNase-free Water to the membrane center of the VNE Column. Stand VNE Column for 2 min.**  
--**Important step!** For effective elution, make sure that the RNase-free Water is dispensed onto the membrane center and is absorbed completely.
- 12. Centrifuge for 2 min to elute the nucleic acid.**
- 12. Store nucleic acid at -70 °C.**