

FT-BY37A0



## Viral DNA/RNA Mini Kit

For different cultured cells supernatant, tissue homogenate and swabs-derived samples

### Product Description

**Catalog #:** BY37A0, 100 rxn  
**Name :** **Viral DNA/RNA Mini Kit - for Amplification and NGS Ready Sample**  
**Description :** The Viral DNA/RNA Mini Kit provides a rapid method for extracting viral DNA/RNA from different cultured cells supernatant, tissue homogenate and swabs-derived samples. The kit is based on silica gel membrane purification technology and does not require  $\beta$ -mercaptoethanol, phenol/chloroform, or any other toxic reagent during the extraction process. The isolated RNA/DNA, characterized by high purity and no residual of impurities, can be applied to reverse transcription, RT-PCR, conventional PCR, NGS and Northern blotting.

<b>Components :</b>	Buffer VL	50 ml	Assist in sample lysis;
	Buffer RW	120 ml	Remove impurities such as proteins and salts;
	RNase-free ddH <sub>2</sub> O	6 ml	Elute DNA/RNA absorbed by the column;
	RNA Columns	100	Specifically absorb DNA/RNA;
	Collection Tubes 2 ml	100	Collect filtrate;
	RNase-free Collection Tubes 1.5 ml	100	Collect DNA/RNA.

**Storage :** Store at 15 ~ 25°C and transport at room temperature.

For Research Use Only

### Workflows



**Sample Lysis:**

Prepare 200  $\mu$ l sample, add 500  $\mu$ l Buffer VL, vortex and mix for 15- 30 sec and centrifuge the mixture to the bottom of centrifuge tubes.

**DNA/RNA Absorption:**

Transfer the mixture to RNA Columns, centrifuge at 12,000 rpm ( $13,400 \times g$ ) for 1 min and discard the filtrate.

**Impurity Removal:**

Add 600  $\mu$ l Buffer RW, centrifuge at 12,000 rpm ( $13,400 \times g$ ) for 30 sec, discard the filtrate, and repeat above steps again. Centrifuge the empty column at 12,000 rpm ( $13,400 \times g$ ) for 2 min.

**DNA/RNA Elution:**

Add 30 - 50  $\mu$ l RNase-free ddH<sub>2</sub>O, keep the column still at room temperature for 1 min and centrifuge the column at 12,000 rpm ( $13,400 \times g$ ) for 1 min.

### Self-Prepared Materials

RNase-free pipette, 1.5 ml RNase-free centrifuge tube, centrifugal machine, vortex oscillator, RNase-free pipette tips



## Notes

1. Equilibrate sample to room temperature before use.
2. The virus may be highly contagious, so some protective measures should be taken.
3. Repeated freeze-thaw should be avoided in case the extracted viral DNA/RNA degrade.
4. Some instruments and consumables should be prepared before use, RNase-free pipette, 1.5 ml RNase-free centrifuge tube, centrifugal machine, vortex oscillator, RNase-free pipette tips.
5. When using this kit, please wear laboratory coat, disposable latex gloves, disposable mask and use RNase-free consumables to prevent RNase contamination.
6. All steps should be performed at room temperature unless otherwise stated.

## Protocol

The following steps should be performed inside the laboratory biosafety cabinet.

1. Add 200µl sample to RNase-free centrifuge tube (complement using PBS or 0.9% NaCl when the sample volume is less than 200ul), then add 500 µl Buffer VL, vortex and mix thoroughly for 15 - 30 sec, and centrifuge the mixture to the bottom of centrifuge tubes.
2. Put RNA Columns inside the Collection Tubes 2 ml, transfer the mixture of Step 1 to RNA Columns, centrifuge the columns at 12,000 rpm (13,400 × g) for 1 min and discard the filtrate.
3. Add 600 µl Buffer RW inside the RNA Columns, centrifuge at 12,000 rpm (13,400 × g) for 30 sec and discard the filtrate.
4. Repeat the Step 3.
5. Centrifuge the empty columns at 12,000 rpm (13,400 × g) for 2 min.
6. Transfer RNA Columns of Step 5 inside another RNase-free Collection Tubes 1.5 ml (Provided in the kit) meticulously, add 30 - 50 µl RNase-free ddH<sub>2</sub>O to the RNA Columns, keep still for 1 min and then centrifuge at 12,000 rpm (13,400 × g) for 1 min.
7. Discard the RNA Columns, and the harvested DNA/RNA can be applied to subsequent assays instantly or kept at -30 ~ -15°C for short-term storage or -85 ~ -65°C for long-term storage.

## FAQ & Trouble Shooting

### ◇ Absorption Columns Clogged

#### ← Excessive impurities

Prepare cell-free samples or obtain the supernatant of cells for subsequent assays

### ◇ No DNA/RNA or very low yield

#### ← The samples underwent repeat freeze-thaw

Use fresh samples and avoid freeze-thaw

#### ← DNA/RNA content of samples was very low

Add some reagents to facilitate the precipitation of DNA/RNA

#### ← The elution was not thorough

Add RNase-free ddH<sub>2</sub>O to the center of the membrane and reduce the elution volume as appropriate. Preheat the RNase free ddH<sub>2</sub>O to 65°C, and prolong the incubation time or perform secondary elution

#### ← The samples were not equilibrated to room temperature

Equilibrate the samples to room temperature before using the kit

### ◇ Inhibition of downstream assays or low purity of extracted RNA/DNA

#### ← Salt residues

Ensure the twice elution using Buffer RW. Additionally, add Buffer RW to the sides of the adsorption column or close the lid of the column and invert 2 to 3 times after adding Buffer RW, which can help to completely wash away any salts on the sides of the column.

#### ← Ethanol residues

Perform the Step 5 and then keep the empty column still at room temperature for 5 min

## Ordering information

Catalog size quantities and prices may be found at <https://www.interchim.com>.

Please inquire for higher quantities (availability, shipment conditions).

Please contact InterBioTech – Interchim for any other information

Hotline : +33(0)4 70 03 73 06 – biosciences@advion-interchim.com

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