



Product data sheet

Interchim Innovations

Interbiochrom - Biochromatography

UptiTip™ Packed WGA

*Sample Preparation Extraction (SPE) of micro quantities.
Immunoglobulines Purification*

Catalog # : BJ3780 UptiTip™ Packed WGA 1-10 µl
 BJ3790 UptiTip™ Packed WGA 10-200 µl

Applications : Affinity isolation of glycoproteins or glycoconjugates by using WGA UptiTip. The slit in the bottom, ~ 1-2 µm wide, permits fluids to pass but not the 50-µm packing material. Thus, no filter is necessary. This permits the purification of glycoconjugates containing terminal N-acetyl glucosyl residues or sialic acid in minimal volumes and minimizes the potential for sample loss. Each tip contains agarose beads with 40 µg (UptiTip™ Packed WGA 1-10 µl) or 100 µg WGA (UptiTip™ Packed WGA 10-200 µl) and sample volumes in the range 1-20 µl (10-200 µl respectively) or larger.

Directions for use

Conditioning Procedure

- a) Tap the UptiTip gently to displace any packing material sticking to the top white cap.
Remove the white caps from top and bottom.
- b) Via a pipette tip inserted in the top of the UptiTip, add 10 µl (50 µl) of loading buffer in order to wet the packing material. Attach the UptiTip to a pipettor or syringe and apply air pressure to force the solution through the packed bed. Remove the UptiTip from the pipettor and repeat this washing procedure 3x.

Suitable solutions are:

- i. Loading Buffer : 100mM sodium phosphate pH 7.0, 0.2M NaCl.
- ii. Wash buffer : 50mM sodium phosphate pH 7.0, 0.2M NaCl
- iii. Elution Buffer : 0.3M Mannose or glucose or N-acety-D-glucosamine in 50mM sodium phosphate pH 7.0, 0.2M NaCl.

NOTE : Do not aspirate (suck up) the liquid. Since there is no filter on top, this will disrupt the packed bed and the material may be sucked into the pipettor. Liquids should always be forced through the packed bed either via positive air pressure or in a microcentrifuge.

Sample Binding

Apply the sample solution as above, attach the UptiTip to a pipettor or syringe, and force the liquid through the packed bed.

NOTE: If this process is slow, then hold the UptiTip onto the pipettor or syringe with one hand and push the plunger slowly ; otherwise, the tip could pop off due to the high pressure.

Sample Washing

Wash the packed bed 2-3x with 10 µl (50 µl) volumes of the binding solution in order to elute salts and other nonretained components.

Sample Release

Wash the packed bed with 2-10 µl (25-50 µl) of releasing solution (bed volume is 8 µl (25 µl)). Repeat and combine the eluents in order to elute all of the adsorbed glycoconjugates.

Protocol of use

A sample can be loaded only from the top of the tip. Please don't try to pull the sample from the bottom of the tip. It works only in one direction.

SpinColumn-in-a-Tip : UptiTips can also be used as a Spin column-in-a-Tip by using the centrifuge adapters. Tips should be centrifuged at speeds between 2000-5000 rpm. The sample will be ready in a few minutes.

- a) The UptiTip™ should be gently tapped so that the dry material does not stick to the white cap.
- b) Remove the white cap from top and bottom of the UptiTip.
- c) Place 50 µl of 70% Isopropanol or 70% Acetonitrile solution to wet the chromatographic material. Addition of 0.05% of TFA or other salt or ion-pairing salts can help in the binding of some protein or peptides for your specific application. **(Please do not pull the sample from the lower end of UptiTip™ as it will disturb the chromatographic media bed. Further more do not interrupt the pushing of the sample in between, as it will cause back pressure and the chromatography media may be sucked into the pipette and disturb the separation. Therefore once you start to press the sample, press all the way to the end until the entire sample has passed through the chromatographic bed. In case you have to interrupt the pressing, first remove the UptiTip™ from the pipette or syringe and then release the pressure of the pipette or syringe).**
- d) Wash the chromatographic media bed volume 2-3 times with the solution of your choice.
- e) Place the sample in the UptiTip and push with the micropipette. In case the solution passes very slowly through the UptiTip, use the syringe supplied with the UptiTips. In that case, hold the tip with one hand firmly and push the syringe slowly, otherwise the tip may pop out due to the pressure and you may lose your sample.
- f) Wash the sample to remove salt or any undesired molecule with water or buffer or other solution for your specific applications.
- g) Elute the sample with the elution buffer.

NOTE :

- a) **The UptiTip does not contain any filters therefore add sample very carefully so that the upper layer of chromatographic material bed is not disturbed.**
- b) **Once you start pressing the pipette to enable sample to pass through the chromatographic bed, don't relieve the pressure in between. First remove the UptiTip from the pipette and then release the press button . If this is not done carefully, the chromatographic bed can be pulled into the pipette and will result in a loss of the sample or the chromatographic bed. This will also disturb the separation quality.**
- c) **Small air bubbles in the bed do not disturb the separation.**
- d) **At times, some peptides or proteins may be absorbed at the filter, however most of time they elute with the elution solution and do not stick to the filter.**

CONCENTRATION OF SAMPLES :

The UptiTip is prepared as above. The dilute solution of biomolecules is passed with the help of the supplied syringe. The syringe supplied is 2.5ml.

The sample should be pushed slowly and gradually and not more than a couple of hundred microliters per minute. A faster flow can also be achieved, if binding affinity of the biomolecule is high towards the chromatographic materials.

Once the biomolecule is bound to the chromatographic material, wash with water or buffer and elute the molecule in a very small volume. By this method, large volumes of several hundred ml samples can be concentrated to few micro liters.

For in vitro R&D use only

Please contact Uptima – Interchim for any other information