

INTENDED USE

The **Chicken anti-BSA IgG** ELISA Kit is an immunoassay suitable for quantifying or titrating total antibody activity specific for bovine serum albumin (BSA) in serum or plasma. Other biological fluids, including tissue culture medium, may be validated for use. For research use only (RUO), not for diagnosis, cure or prevention of the disease.

GENERAL INFORMATION

Bovine serum albumin (BSA) is the major protein of bovine serum (also fetal calf serum), used as a required nutrient for the *in vitro* culture of many cell types, including cell lines for production of recombinant proteins used as pharmaceutical drugs. Such recombinant protein culture products are purified from culture medium components, including BSA, which even as minor contaminants may affect drug efficacy and side effects. BSA is also highly immunogenic in mammals and can, therefore, elicit anti-BSA antibodies in the host receiving the BSA-contaminated drug. The measurement of residual culture components such as BSA and the immunological host response, i.e., anti-BSA antibodies, is a requirement of the manufacturing processes of recombinant protein drug production.

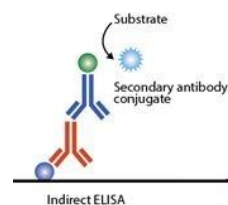
Purified BSA is also used as a carrier protein for chemical attachment of small molecules, including peptides and other haptens, to produce an effective vaccine for production of antibodies specific for the attached molecule. The BSA-hapten immunization effectively produces anti-BSA antibodies concomitant with anti-hapten antibodies. These 'by-stander' antibodies, which may interfere with the subsequent use and interpretation of the anti-hapten activity, are commonly removed by solid phase BSA adsorbents.

The ADI Chicken Anti-BSA IgG ELISA is a sensitive and convenient method for detecting and quantifying anti-BSA IgG antibodies arising from BSA contamination of administered drugs, and for following anti-BSA antibodies from carrier-hapten immunizations.

PRINCIPLE OF THE TEST

The **Chicken anti-BSA IgG** ELISA kit is based on the binding of chicken anti-BSA IgG in samples to BSA immobilized on the microwells, and anti-BSA IgG antibody is detected by anti-chicken IgG-HRP conjugate. After a washing step, chromogenic substrate (TMB) is added and color (blue) is developed, which is

directly proportional to the amount of anti-BSA present in the sample. Stopping Solution is added to terminate the reaction (converts blue to yellow color), and A450nm is then measured using an ELISA microwell reader. The activity of anti-chicken IgG antibody in samples is calculated relative to anti-BSA calibrators.



PRODUCT SPECIFICATIONS

Specificity

Purified BSA is used to coat the microwells; thus the assay is specific for antibodies directed to BSA. The anti-Chicken IgG HRP conjugate reacts specifically with chicken IgG antibodies that bind to BSA on the plate; IgA, IgM and IgE antibody would not be measured above background signals.

KIT CONTENTS

The microtiter well plate and all other reagents, if unopened, are stable at 2-8°C until the expiration date printed on the box label. Stabilities of the working solutions are indicated under Reagent Preparation.

To Be Reconstituted: Store as indicated.

Component	Preparation Instructions
Wash Solution Concentrate (100x) Cat. No. WB-100, 10ml	Dilute the entire volume 10ml + 990ml with distilled or deionized water into a clean stock bottle. Label as Working Wash Solution and store at ambient temperature until kit is used entirely.
Sample Diluent Concentrate (20x) Cat. No. SD-20B, 10ml	Dilute the entire volume, 10ml + 190ml with distilled or deionized water into a clean stock bottle. Label as Working Sample Diluent and store at 2-8°C until the kit lot expires or is used up.
Anti-Chicken IgG-HRP Conjugate Concentrate (100x) Part: H-ChG.2a11, 0.15ml	Peroxidase conjugated anti-Chicken IgG in buffer with detergents and antimicrobial as stabilizers. Dilute fresh as needed; 10ul of concentrate to 1ml of Working Sample Diluent is sufficient for 1 8-well strip. Use within the working day and discard. Return 100X to 2-8°C storage.

Ready For Use: Store as indicated on labels.

Component	Part	Amt	Contents
BSA Microwell Strip Plate	8001	8-well strips (12)	Coated with BSA, and post-coated with stabilizers.
Anti-BSA Calibrators			
10 U/ml	710-112B	0.65 ml	Four (4) vials, each containing anti-BSA antibody in buffer with antimicrobial.
25 U/ml	710-112C	0.65 ml	
50 U/ml	710-112D	0.65 ml	
100 U/ml	710-112E	0.65 ml	
Anti-BSA Positive Control	710-112PC	0.65ml	Anti-BSA antibody, diluted in buffer with antimicrobial. [value range on vial]
TMB Substrate	80091	12 ml	Chromogenic substrate for HRP containing TMB and peroxide.
Stop Solution	80101	12 ml	Dilute sulfuric acid.

Materials Required But Not Provided:

- Pipettors and pipettes that deliver 100ul and 1-10ml. A multi-channel pipettor is recommended.
- Disposable glass or plastic 5-15ml tubes for diluting samples and Anti-Chicken IgG HRP Concentrate.
- Graduated cylinder to dilute Wash Concentrate; 0.2 to 1L.
- Distilled or deionized water to dilute reagent concentrates.
- Microwell plate reader at 450 nm wavelength.

ASSAY DESIGN AND SET-UP

CAUTION: Avoid solutions or containers with BSA, which would block or diminish anti-BSA activity in samples.

Sample Collection and Handling

Culture medium (BSA-free), serum and other biological fluids may be used as samples with proper dilution to avoid solution matrix interference. For **serum**, collect blood by venipuncture, allow clotting, and separate the serum by centrifugation at room temperature. For other samples, including **tissue culture media**, clarify the sample by centrifugation and/or filtration prior to dilution in Sample Diluent. If samples will not be assayed immediately, store refrigerated for up to a few weeks, or frozen for long-term storage. Avoid freeze-thaw cycles.

Assay Design

Review Calculation of Results (p5-7) and Limits of the Assay (above) before proceeding:

- Select the proper sample dilutions accounting for expected potency of positives and minimizing non-specific binding (NSB) and other matrix effects; for example, net signal for non-immune samples should be <0.5 OD. This is usually 1/100 or greater dilution for chicken sera with normal levels of IgG and IgM.
- Run a Sample Diluent **Blank**. This signal is an indicator of proper assay performance, especially of washing efficacy, and is used for net OD calculations, if required (**See Method A,B**). Blank OD should be <0.3.
- Run a set of **Calibrators**. Calibrators validate that the assay was performed to specifications; results can be used to normalize between-assay variation for enhanced precision. Reading values off a Calibrator curve, has limitations. See Limits of the Assay (page 7).
- Run a range of sample dilutions for expected higher positives that allows calculation of antibody **Titer** (when specific titer is at least 4-fold higher than non-immune). **See Method C**.
- Run samples in duplicate if used for quantitation; non-immunes that are significantly lower than immunes may be run in singlicate. The Calibrators that are used for quantitation, e.g., for between-assay normalization, should be run in duplicate. When determining titer from a dilution curve, singlicates can be run if more than two dilution points are used for titer calculations.

Plate Set-up

Bring all reagents to room temperature (18-30° C) equilibration (at least 30 minutes).

- Determine the number of wells for the assay run. Duplicates are recommended, including 8 Calibrator wells and 2 wells for each sample and control to be assayed.
- Remove the appropriate number of microwell strips from the pouch and return unused strips to the pouch. Reseal the pouch and store refrigerated.
- Add 200-300ul Working Wash Solution to each well and let stand for about 5 minutes. Aspirate or dump the liquid and pat dry on a paper towel before sample addition.

Assay Procedure

ALL STEPS ARE PERFORMED AT ROOM TEMPERATURE. After each reagent addition, gently tap the plate to mix the well contents prior to beginning incubation.

1. 1st Incubation [100ul – 60 min; 4 washes]

- Add 100ul of calibrators, samples and controls each to pre-determined wells.
- Tap the plate gently to mix reagents and incubate for 60 minutes.
- Wash wells 4 times and pat dry on fresh paper towels. As an alternative, an automatic plate washer may be used. Improper washes may lead to falsely elevated signals and poor reproducibility.

2. 2nd Incubation [100ul – 30 min; 5 washes]

- Add 100ul of diluted Anti-Chicken IgG HRP to each well.
- Incubate for 30 minutes.
- Wash wells 5 times as in step 2.

3. Substrate Incubation [100ul – 15 min]

- Add 100ul TMB Substrate to each well. The liquid in the wells will begin to turn blue.
- Incubate for 15 minutes in the dark, e.g., place in a drawer or closet.

Note: If your microplate reader does not register optical density (OD) above 2.0, incubate for less time, or read OD at 405-410 nm (results are valid).

4. Stop Step [Stop: 100ul]

- Add 100ul of Stop Solution to each well.
- Tap gently to mix. The enzyme reaction will stop; liquid in the wells will turn yellow.

5. Absorbance Reading

- Use any commercially available microplate reader capable of reading at 450nm wavelength. Use a program suitable for obtaining OD readings, and data calculations if available.
- Read absorbance of the entire plate at 450nm using a single wavelength within 30 minutes after Stop Solution addition. If available, program to subtract OD at 630nm to normalize well background.



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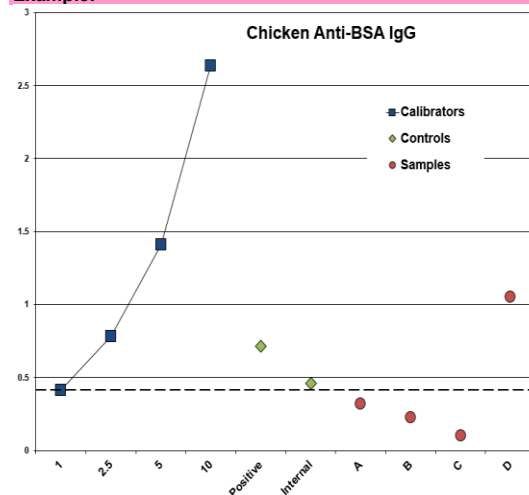
INTERPRETATION OF RESULTS

Method A. Antibody Activity Threshold Index

Compare Samples to **10 U/ml Calibrator** or **Internal Control**

= **Positive/Negative Cut-off.**

Example:



Results

The **sensitivity** of the assay to detect anti-BSA IgG, from either natural exposure or vaccination, is controlled so that the **10 U/ml Calibrator** represents a threshold OD for most true positives in chicken serum diluted to 1:100 or greater. Visual inspection of the data in the above graph shows the following:

Calibrators – dilution curve of an anti-BSA antibody, derived from BSA immunization, shows the OD range of the assay; high value indicates optimal sensitivity of the assay.

10 U/ml: a 'Cut-off' line has been drawn to indicate a threshold distinguishing between **Positive/Negative**. This is not a clear-cut threshold, rather a low OD area that could represent either low positives or high background negatives.

Internal Control – a low level positive from an immunized animal that represents the lab's experience in distinguishing low positive from negative samples (not supplied). This should be run in each assay to supplement the 10 U/ml Calibrator for Positive/Negative discrimination purposes.

Samples A,B,C,D – 3 samples (A, B, C) are negative: below the threshold; 1 sample (D) is positive: clearly above the threshold.

The 10 U/ml Calibrator can be used to calculate a **Threshold Index** that numerically discriminates Positive/Negative:

- ❖ Divide each Sample net OD by the 10 U/ml Calibrator net OD. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

INTERPRETATION OF RESULTS (cont)

Method B. Positive Index

Experimental sample values may be expressed relative to the values of Control or Non-immune samples, by calculation of a **Positive Index**. One typical method is as follows:

1. Calculate the net OD mean + 2 SD of the Control/Non-immune samples = **Positive Index**.
2. Divide each sample net OD by the Positive Index. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

A sample value would be **Positive** if significantly above the value of the pre-immune serum sample or a suitably determined non-immune panel or pool of samples, tested at the same sample dilution.

This calculation also **quantifies** the positive Antibody Activity level, assigning a higher value to samples with higher Antibody Activity, and vice versa.

Example:

Sample	Assay Net OD		Calculated Antibody Activity	
	Control	Exptl	Control	Exptl
1	0.244	C 2.293	0.57	5.34
2	0.204	C 1.490	0.48	3.47
3	0.237	C 0.833	0.55	1.94
4	0.26	C 0.326	0.61	0.76
5	0.388	P 1.106	0.90	2.58
6	0.407	I 0.310	0.95	0.72
7	0.288	E 0.672	0.67	1.56
8	0.263	E 0.363	0.61	0.85
9	0.322	E 0.560	0.75	1.31
10	0.343	E 0.490	0.80	1.14
Mean	0.295			
SD	0.067			
Mean +2 SD	0.429	= Positive Index		

Results

Experimental Samples are represented as follows:

C – Calibrator
P – Positive Control
I – Internal Control; lab's threshold positive serum
E – Experimental sample

ASSAY PERFORMANCE

Method C. Titers from Sample Dilution Curves

The titer of antibody activity calculated from a dilution curve of each sample is recommended as the most accurate quantitative method. Best precision can be obtained using the following guidelines:

1. Use an OD value Index in the mid-range of the assay (2.0 – 0.5 OD); this provides the best sensitivity and reproducibility for comparing experimental groups and replicates. An arbitrary 1.0 OD is commonly used.
2. Prepare serial dilutions of each sample to provide a series that will produce signals higher and lower than the selected index. With accurate diluting, duplicates may not be required if at least 4 dilutions are run per sample.
3. A 5-fold dilution scheme is useful to efficiently cover a wide range which produces ODs both above and below 1.0 OD. The dilution scheme can be tightened to 3-fold or 2-fold for more precise comparative data.
4. The Positive and Sensitivity Control values can be used to normalize inter-assay values.

Calculations

1. On a log scale of inverse of Sample Dilution as the x-axis, plot the OD values of the two dilutions of each positive sample having ODs above and below the OD value of the Index (arbitrary or selected Calibrator).
2. From a point-to-point line drawn between the two sample ODs, read the dilution value (x-axis) corresponding to the OD of the selected Index
 = **IgG Antibody Activity Units**

Limits of the Assay

- To quantitate antibody activity from a calibrator curve (such as provided with the kit), the dilution curve of the samples must be parallel to the calibrator curve, to avoid different values being obtained from different regions of the curve. Antibodies that are not matched in BSA avidity will often have non-parallel dilution curves. In these cases, antibody activity is best expressed using the other calculation methods.
- The sensitivity of the assay may be increased to perhaps convert a borderline sample to a positive by using a lower dilution of the sample, e.g., 1/50. The values of negatives may increase, so an alternative threshold should be established using known negatives to develop a **Positive Index** (page 6), or by using known **Internal Controls** as discriminator for a **Threshold Control** (instead of the kit 1 U/ml Calibrator Control)

PRECAUTIONS AND SAFETY INSTRUCTIONS

Calibrators, Sample Diluent, and Antibody HRP contain bromonitrodioxane (BND: 0.05%, w/v). Stop Solution contains dilute sulfuric acid. Follow good laboratory practices, and avoid ingestion or contact of any reagent with skin, eyes or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water. MSDS for TMB, sulfuric acid and BND can be requested or obtained from the ADI website.

Chicken Anti-Bovine Serum Albumin (BSA) IgG ELISA Kit

Cat. No. 710-120-BSC, 96 tests

For Quantitation of anti-BSA IgG in Serum, Plasma or other Biological Solutions

For research use only (RUO), not for diagnosis, cure or prevention of the disease.



your distributor



ALPHA DIAGNOSTIC INTERNATIONAL

ELISA Kit Components	Amount	Part
BSA Coated Microwell Strip Plate	8-well strips (12)	8001
Anti-BSA Positive Control	0.65 ml	710-112PC
Anti-BSA Calibrator 10 U/ml	0.65 ml	710-112B
Anti-BSA Calibrator 25 U/ml	0.65 ml	710-112C
Anti-BSA Calibrator 50 U/ml	0.65 ml	710-112D
Anti-BSA Calibrator 100 U/ml	0.65 ml	710-112E
Anti-Chicken IgG HRP Conjugate (100X)	0.15 ml	H-ChG.2a11
Sample Diluent (20x)	10 ml	SD20B
Wash Solution Concentrate (100X)	10 ml	WB-100
TMB Substrate	12 ml	80091
Stop Solution	12 ml	80101
Product Manual	1 ea	M-710-120-BSC