

FT-A2WQP0



## Eosin Methylene Blue Agar

*For the isolation, cultivation and differentiation of gram negative enteric bacilli from clinical and other specimens*

### Product Description

|                         |  |
|-------------------------|--|
| <b>Name :</b>           | <b>Eosin Methylene Blue Agar</b>                                     |
| <b>Catalog Number :</b> | A2WQP0, 500 g  |
| <b>Storage:</b>         | 2-25°C - Once opened keep powdered medium closed to avoid hydration. |

### Directions for use

#### Formula

- |                              |                             |
|------------------------------|-----------------------------|
| • Bacteriological Peptone 10 | • Eosin Y 0.4               |
| • Lactose 5                  | • Methylene Blue 0.065      |
| • Sucrose 5                  | • Bacteriological Agar 13.5 |
| • Dipotassium Phosphate 2    |                             |

Final pH 7.2 ± 0.2 at 25°C

#### Preparation

Suspend 36 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C, mix well, avoiding the formation of bubbles and dispense carefully into Petri Dishes. DO NOT OVERHEAT. The prepared medium should be stored at 8-15°C. The color is tournasol blue. Sterilization reduces the methylene blue, leaving the medium orange in color. The normal purple may be restored by gently mixing. The reduced medium should be shaken to oxidize the methylene blue; otherwise a dark zone from the top extending downwards will gradually appear.

The dehydrated medium should be homogeneous, free flowing and purple-rose flocculent precipitate in color. If there are any physical changes, discard the medium.

#### Uses

EOSIN METHYLENE BLUE AGAR is a differential medium similar to Levine EMB Agar (Cat. 1050) and is used for the isolation of Enterobacteria. The use of Eosin Y and Methylene Blue enable differentiation between lactose-fermenting and non-fermenting organisms. It is widely used in medical bacteriology, in techniques recommended by APHA and for the detection and enumeration of coliforms, contaminants of foods and drinking water. Peptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Sucrose is added to Lactose as a fermentable

#### FT-A2WQP0

carbohydrate to detect coliforms that ferment sucrose more readily than lactose. Eosin Y and Methylene blue dyes are both partial inhibitors of Gram-positive bacteria and pH indicators. Due to the lactose and sucrose, the medium can be differential in primary culture: *Salmonellae* and *Shigellae* which are lactose-negative can be differentiated from other lactose-negative and sucrose-positive organisms such as *Proteus vulgaris*, *Citrobacter* and *Aeromonas*. Dipotassium phosphate acts as a buffer system and Bacteriological agar is the solidifying agent.

For the isolation of enteric pathogens from clinical samples, inoculate onto a small area of one quadrant of EMB Agar and streak for isolation, allowing discrete colonies to develop. Incubate at  $35 \pm 2^\circ\text{C}$  and observe at 24 hours and again at 48 hours. *Salmonella* and *Shigella* colonies are translucent and amber colored or colorless. Coliforms that use lactose and/or sucrose produce blue-black colonies with dark centers and a greenish metallic sheen. Other coliforms such as *Enterobacter* form mucoid, pink colonies. Strains of *Enterococcus faecalis* are partially inhibited on this medium and appear as colorless colonies. As the medium is moderately inhibitory some staphylococci, streptococci and yeast may grow. Also some Gram-negative non-fermenting bacilli may appear as non-lactose fermenters. Further Biochemical tests are necessary for genus identification.

#### Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubation at a temperature of  $35 \pm 2^\circ\text{C}$  and observed after 24-48 hours.

| Microorganisms                           | Growth    | Colony Color              |
|--|-----------|---------------------------|
| <i>Enterobacter aerogenes</i> ATCC 13048 | Good      | Pink                      |
| <i>Escherichia coli</i> ATCC 25922       | Good      | Green with metallic shine |
| <i>Salmonella typhimurium</i> ATCC 14028 | Good      | Colorless                 |
| <i>Pseudomonas aeruginosa</i> ATCC 10145 | Good      | Colorless                 |
| <i>Staphylococcus aureus</i> ATCC 25923  | Inhibited | Colorless                 |

#### References

American Public Health Association. Diagnostic Procedures and Reagents. 2<sup>nd</sup> Ed. APHA, Inc. New York, 1950  
 A.P.H.A Examination of dairy products. 10<sup>th</sup> Ed. APHA, Inc. New York, 1953. Society of American Bacteriologists.  
 Manual of Microbiological Methods MacGraw-Hill New York, 1957.

### Ordering information

Catalog size quantities and prices may be found at <http://www.interchim.com>.  
 Please inquire for higher quantities (availability, shipment conditions).

For any information, please ask : Uptima / Interchim; Hotline : +33(0)4 70 03 73 06

[Order on-line](#) or [Contact](#) your local distributor

**Disclaimer :** Materials from Uptima are sold **for research use only**, and are not intended for food, drug, household, or cosmetic uses.  
 Uptima is not liable for any damage resulting from handling or contact with this product.